



## BENEFICIAL IMPACT OF DIPYRIDAMOLE AND DIDS ON ERYTHROCYTE MEMBRANE AT MEMBRANOPATHY

Iv. Ivanov\*, B. Paarvanova

Department of Physics, Biophysics, Roentgenology and Radiology, Medical Faculty, Trakia University, Stara Zagora, Bulgaria,

### ABSTRACT

Spectrin and the band 3, major proteins of human erythrocyte membrane, are altered in patients with hereditary spherocytosis (HS). The band 3 is implicated in the permeability transition of erythrocyte membranes (EM) at 61°C ( $T_g$ ), and its monomer is specifically stabilized by DIDS (4,4'-diisothiocyanostilbene-2,2'-disulphonate) and Dipyridamole (2,6-bis(diethanolamino)-4,8-dipiperidinopyrimido[5,4-d] pyrimidine). **PURPOSE.** To study in vitro the impact of DIDS and dipyridamol on the band 3 in anemic patients as revealed by the changes in the  $T_g$  temperature. **METHODS.** We used thermal analysis of the impedance (70 kHz) of erythrocytes suspended in isotonic 30 mM NaCl/mannit media in order to detect permeability transition at  $T_g$  and the heat denaturation of spectrin at 49.5 °C. **RESULTS:** In patients (n = 8) with HS the  $T_g$  transition was detected at 56-57°C and accompanied by severe instability of EM after the spectrin denaturation temperature. DIDS (30 μM, irreversible) and dipyridamole (100 μM, reversible) increased the  $T_g$  to about 60°C and subdued the preceding membrane instability. **CONCLUSION.** Dipiridamole and DIDS both stabilized the structure of EM from patients with HS. Thermal dielectroscopy is useful test for population screening of asymptomatic forms of HS.

**Key words:** band 3, hemolytic anemia, membranopathy, thermal hemolysis.

### INTRODUCTION

The integrity and mechanical properties of human erythrocyte membrane (EM) rely chiefly on the two major membrane proteins, spectrin and the band 3. They are frequently altered in some inherited anemic conditions such as hereditary spherocytosis (HS) (1-3).

Band 3 is an integral protein whose monomers are associated in dimers and larger oligomers. According to microcalorimetric studies the monomer of band 3 has high structural stability as evident by its high temperature of denaturation, 68°C (4). In addition, the structural stability of band 3 monomer is strongly increased by DIDS and dipyridamole. DIDS is a membrane impermeable covalent amino reagent which at micromolar concentrations (<50 μM) specifically binds the dimer of band 3 and step-

wisely increases the denaturation temperature of the monomer by 13°C (5). Similar, although reversible, effect on the stability of band 3 monomer is produced by dipyridamole (6). At anemic conditions caused by hereditary red cell membrane disorders the alteration in band 3 is not accompanied by a reduction in the denaturation temperature of band 3 monomer. The band 3 of EMs is implicated, however, in another milder, predenaturational thermally – induced membrane transition concerning the oligomeric structure of this protein (7). This permeability transition affects the ion permeability of EM and is responsible for thermal hemolysis. In healthy humans the permeability transition is centered at 61±0.2°C, the so called  $T_g$  temperature. Interestingly enough, for a limited group of patients with HS the  $T_g$  temperature had a values lowered by about 4-5°C (8). In the presented study we used larger group of patients with HS and studied the *in vitro* effect of DIDS and dipyridamol on the  $T_g$

\*Correspondence to: Ivan Tanev: Dept. Physics, Biophysics, Rentgenology and Radiology, Medical Faculty, Thracian University, Armeyska Str. 11, Stara Zagora 6000, Bulgaria. E-mail: [ivanov\\_it@gbg.bg](mailto:ivanov_it@gbg.bg)

temperature of the permeability transition in their EMs.

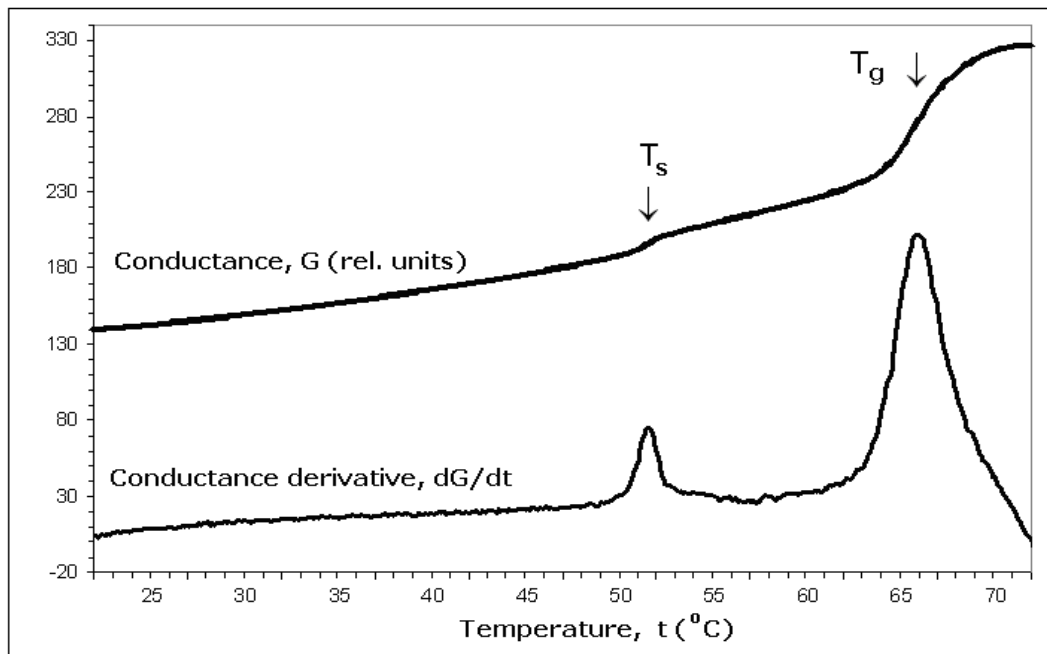
## MATERIALS AND METHODS

### Materials

Dipyridamole (2,6-bis(diethanolamino)-4,8-dipiperidinopyrimido[5,4-d] pyrimidine), DIDS (4,4'-di-isothiocyanostilbene-2,2'-disulphonate), sorbitol and NaCl were all purchased from Sigma, St. Louis, MO.

### Impedance spectroscopy of the heated erythrocyte suspensions

The isolation and DIDS treatment of erythrocytes was described previously (7). The binding of DIDS to band 3 of EMs was carried out according to (9). To determine the permeability transition at  $T_g$  and the heat denaturation of spectrin at 49.5 °C we used thermal analysis of the impedance of erythrocyte suspensions as described previously (7).



**Figure 1.** Temperature profile of the conductance,  $G$ , and temperature-derivative of conductance,  $dG/dt$  of erythrocytes suspended in isotonic 30 mM NaCl/mannit solution. The hematocrit, heating rate and frequency were 0.30, 2.5 °C/min and 70 kHz, respectively.

## RESULTS

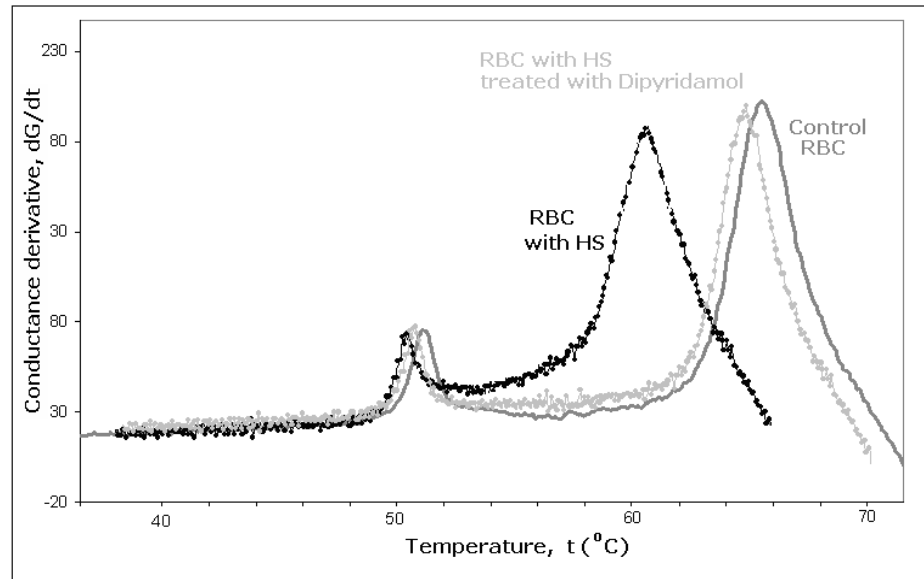
Temperature-induced variations in the conductance,  $G$ , of heated erythrocyte suspensions are shown in **Figure 1**. Superimposed over the continuous Boltzmann type temperature dependence of  $G$  two sharp, threshold changes were registered with mid-point temperatures  $T_s$  and  $T_g$ , respectively. These changes are better identifiable as sharp peaks on the derivative thermogram of suspension conductance. The two top temperatures,  $T_s$  and  $T_g$ , slightly depended on the heating rate applied and at heating rates extrapolated to zero their values were 49.5°C and 61.0°C, respectively. During repeated measurements of samples, taken from a same blood, the  $T_s$  and  $T_g$  temperatures displayed variations by  $\pm 0.2$  °C from respective mean values.

Previous studies evidenced that the peaks centered at the  $T_s$  and  $T_g$  top temperatures both correspond to thermally-induced structural transitions in the plasma membrane of erythrocytes (10). The change in the suspension conductance at  $T_s$  is explained by the effect of spectrin denaturation on the dielectric polarization of EM (11). According to microcalorimetric study (12) the heat denaturation of spectrin takes place at 49.5°C coinciding with  $T_s$ . The peak centered at  $T_g$  demonstrates the activated passive diffusion of cytosole ions due to membrane transition involving a mild, pre-denaturational alteration in the band 3 (7).

Compared to control ones, the  $T_g$  temperature was lowered by about 4 to 5°C for the membranes of erythrocytes with HS (**Figure 2**). Similar outcome was previously reported for a

smaller ( $n = 2$ ) group of patients with HS (8). This finding is in line with the report (13) that ion permeability of erythrocytes with HS is

increased. It possibly represents a distinctive feature of the erythrocytes from patients with HS and hereditary altered band 3.



**Figure 2.** Temperature profile of  $dG/dt$  of erythrocyte suspensions. Dark gray curve - control erythrocytes, black curve - erythrocytes with moderate form of HS. Gray curve demonstrates the effect produced by Dipyridamol (100  $\mu\text{M}$ ) and DIDS (30  $\mu\text{M}$ ) on erythrocytes with a moderate form of HS ( $n = 4$ ). Other details as for **Figure 1**.

The *in vitro* treatment of such erythrocytes with DIDS (30  $\mu\text{M}$ ) significantly increased their  $T_g$  temperature close to that in control cells (**Figure 2**). This result is in concert with the report that the band 3 is important participant in the permeability transition in erythrocyte membrane at  $T_g$  (7). At micromolar concentrations (50–100  $\mu\text{M}$ ) the medicine dipyridamole (persantine, antistenocardine) also increased the  $T_g$  temperature. In contrast to DIDS, the effect of dipyridamole was reversible and concentration dependent (not shown).

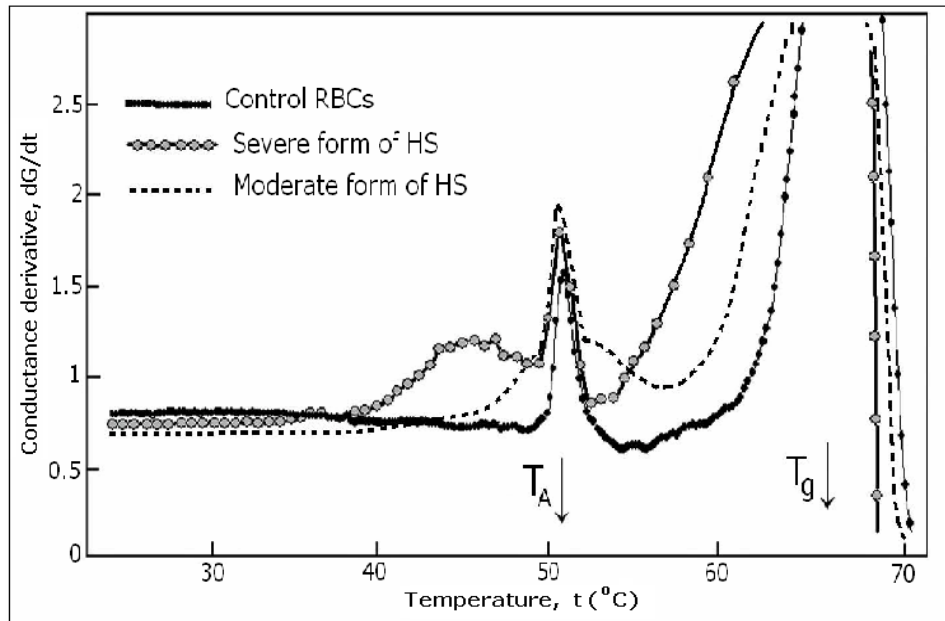
Except the reduction of  $T_g$  temperature, an additional membrane instability about the  $T_s$  temperature was apparent on the  $dG/dt$  curve of erythrocytes with HS (**Figure 3**). This membrane instability was demonstrated by a third peak with much broader half-height width than the spectrin denaturation peak at  $T_s$ . The amplitude of this peak was higher and its top temperature was lower in patients with severe form of HS while in patients with moderate condition this peak was poorly distinguishable (**Figure 3**). DIDS (30  $\mu\text{M}$ ) and dipyridamole (50–100  $\mu\text{M}$ ) both reduced significantly the amplitude of the third peak (not shown). Similar to the spectrin denaturation peak at  $T_s$  (11), the third peak had

dispersive character as its amplitude was reduced with the frequency and it completely vanished above 200 kHz (not shown).

## DISCUSSION

This study, combined with previous one (8), reports a 4–5°C reduction in the  $T_g$  temperature in erythrocytes of patients with HS. This could be explained taking into account that the two events, the permeability transition at  $T_g$  and thermal denaturation at 68°C, concern different types of band 3 structure. While the first event increase the oligomerization of band 3 (7) the latter event destroys the secondary and tertiary structure of this protein. Since the same erythroid isoform of band 3 is expressed in the acid secreting cells of kidney the genetically altered band 3 could also affect the regulation of urine acidity, according to (15).

The band 3 is phylogenetically preserved in most cells of vertebrate animals (14). Erythroid band 3 mutations cause 15–25% of cases of HS, and 100% of cases of southeast Asian ovalocytosis (1). HS affects 1 in 2000 people of Northern European ancestry and at least 1 in 5000 in general (1, 2, 16). Hence, the applied in this study method could be used for broad population screening.



**Figure 3.** Temperature profile of  $dG/dt$  for erythrocytes with HS ( $n = 4$ ) displaying varying severity, compared to control ones. Other details as for **Figure 1**.

## CONCLUSION

In contrast to microcalorimetry, the thermal analysis of suspension impedance detected the inherited red cell membrane disorder in band 3 protein of anemic erythrocytes. This was evident by the significant (4-5°C) decrease in the  $T_g$  temperature of the permeability transition implicating a mild change in band 3. This reduction in  $T_g$  temperature was largely compensated by dipyrindamol and DIDS, two chemicals known to stabilize the structure of band 3.

## REFERENCES

1. An, X. and Mohandas, N., Disorders of red cell membrane. *Br J Haematol*, 141: 367-375, 2008.
2. Barcellini, W., Bianchi, P., Fermo, E., Imperiali, F.G., Marcello, A.P., Vercellati, C., Zaninoni, A. and Zanella, A., Hereditary red cell membrane defects: Diagnostic and clinical aspects. *Blood Transfusion*, July, 9(3):274-277, 2011.
3. Gordon-Smith, E.C. and Mohandas, N., Hereditary disorders of the red cell membrane. In: Green AR, Hoffbrand AV, Catovsky D et al (eds). *Postgraduate Haematology*. Oxford, Wiley-Blackwell, 2010.
4. Maneri, L.R. and Low, P.S., Structural stability of the erythrocyte membrane anion transporter, band 3, in different lipid environment. *J Biol Chem*, 263:16170-16178, 1988.
5. Snow, J.W., Brandts, J.F., and Low, P.S., The effects of anion transport inhibitors on structural transitions in erythrocyte membranes. *Biochim. Biophys. Acta*, 512:579-591, 1978.
6. Legrum, B. and Passow, H., Inhibition of inorganic anion transport across the human red blood cell membrane by chloride-dependent association of dipyrindamol with a stilbene disulfonate binding site on the band 3 protein. *Biochimica et Biophysica Acta*, 979(2):193-207, 1989.
7. Ivanov, I.T., Zheleva, A. and Zlatanov, I., Anion exchanger and the resistance against thermal hemolysis. *International Journal of Hyperthermia*, 27 (3):286-296, 2011.
8. Ivanov, I.T., Tolekova, A. and Chakaarova P., Erythrocyte membrane defects in hemolytic anemias found through derivative thermal analysis of electric impedance. *J. Biochem. Biophys. Methods*, 70:641-648, 2007.
9. Macey, R., Adorante, J.S. and Orme, F.W., Erythrocyte membrane potentials determined by hydrogen ion distribution. *Biochim Biophys Acta*, 512:284-295, 1978.
10. Ivanov, I.T., Allometric dependence of the life span of mammal erythrocytes on thermal

- stability and sphingomyelin content of plasma membranes. *Comparative Biochemistry and Physiology, Part A*, 147:876-884, 2007.
11. Ivanov, I.T., Paarvanova, B. and Slavov, T., Dipole relaxation in erythrocyte membrane: Involvement of spectrin skeleton. *Bioelectrochemistry* 88:148–155, 2012.
  12. Brandts, J.F., Erickson, L., Lysko, K., Schwartz, A.T. and Taverna, R.D., Calorimetric studies of the structural transitions of the human erythrocyte membrane. The involvement of spectrin in the A transition. *Biochemistry*, 16:3450-3454, 1977.
  13. Bruce, L. J., Hereditary stomatocytosis and cation leaky red cells - Recent developments. *IVANOV IV., et al. Blood Cells, Molecules, and Diseases*, 42:216–222, 2009.
  14. Alper, S.L., The band 3-related anion exchanger (AE) gene family. *Annu Rev Physiol*, 53:549–564, 1991.
  15. Rysava, R., Tesar, V., Jirsa Jr, M., Brabec, V. and Jarolim, P., Incomplete distal renal tubular acidosis coinherited with a mutation in the band 3 (AE1) gene. *Nephrol Dial Transplant*;12(9):1869–1873, 1997.
  16. Fauci, A., Braunwald, E., Kasper, D., Hauser, S., Longo, D., Jameson, L. and Loscalzo, J., Harrison's principles of internal medicine (17<sup>th</sup> ed.), New York: *McGraw-Hill Medical*. pp. Chapter 106, 2008.